

A new method to use benthic chlorophyll *a* for water quality assessment

Brad Rogers, PhD candidate, Environmental Science

Dan Storm, Professor, Biosystems Engineering

Derek West, Undergraduate Student, Biosystems Engineering

Andy Dzialowski, Associate Professor, Zoology

Bill Henley, Professor, Botany

Oklahoma State University

Stillwater, Oklahoma

Support Provided by:

Oklahoma Conservation Commission

US Environmental Protection Agency, Region VI

OSU Environmental Sciences Graduate Program

OSU Biosystems and Agricultural Engineering

Background

- **Environmental monitoring**
 - **EMAP (environmental monitoring and assessment program)**
 - **EPA wanted more comprehensive monitoring**
 - **Statistical comparisons**
 - **Long term trends**
 - **Prediction**
 - **Data storage**



- **Nonpoint source management program (Sec 319)**
 - **Water quality**
 - **Stream habitat**
 - **Aquatic communities**

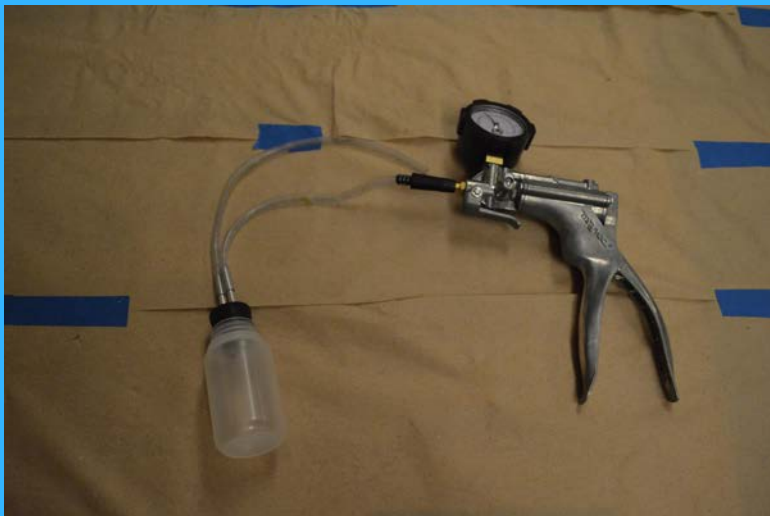
Biologic Monitoring

- **Assess ecologic conditions**
 - Includes adjacent land use/land cover
 - In Stream Habitat
 - Aquatic communities
 - Fish
 - Macrophytes
 - Periphyton



Tar Creek (Miami OK)

Periphyton Sampling Traditional Method



Periphyton Sampling

BenthoTorch®

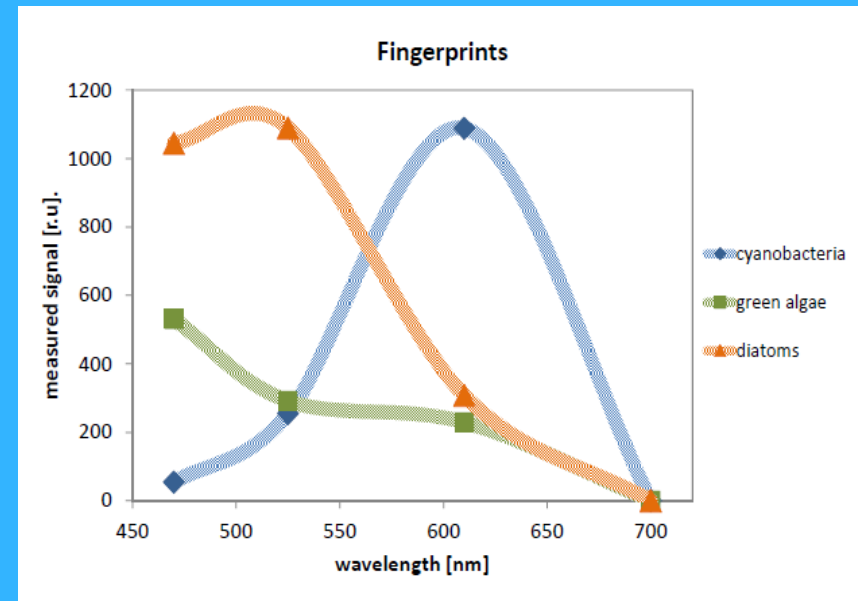
- **Commercial Fluorometer**
 - *In situ* and *in vivo*
 - Provides results in 20 seconds
 - Stores files for later upload
 - Measure total biomass (by chlorophyll fluorescence)
 - Internal proprietary algorithm for relative abundance for three periphyton divisions
 - Used in monitoring and research across the world
 - Two published comparison with traditional method



BenthoTorch®

Based on Principal of Fluorescence

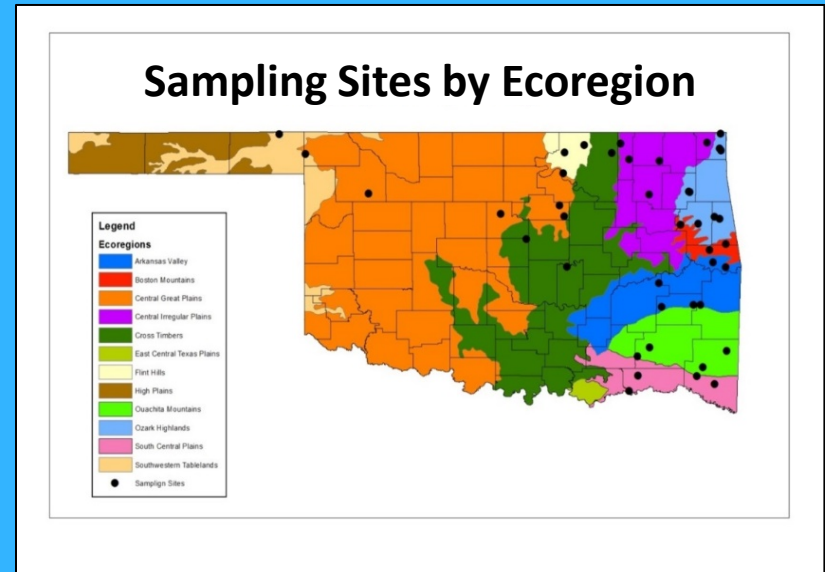
- LED wave lengths (nm): 470, 525, 610, 700
- Light directed at algae in pulses (PAM)
- Chlorophyll emits light at longer wavelength (Stokes Shift)
- 700 nm used to compensate for background reflection



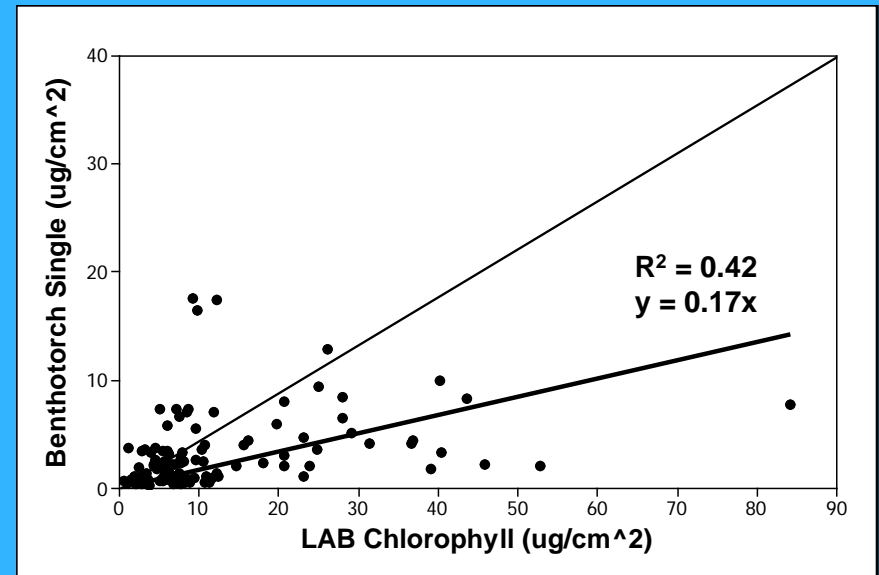
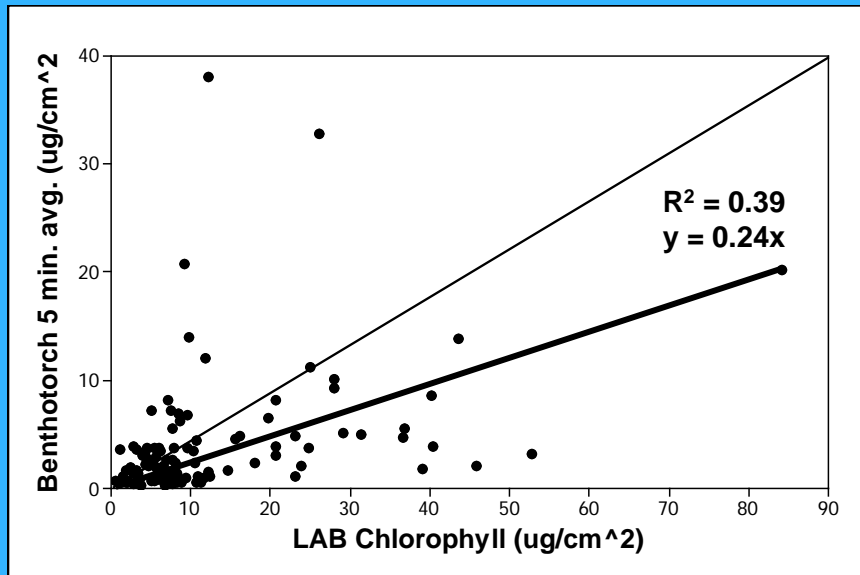
BenthoTorch®®
Algal Class Fingerprint

Objective

- Assess the accuracy of the Benthotorch[®] compared to traditional methods in estimating total benthic algal biomass across major stream types and conditions throughout Oklahoma



Results: Field 2014



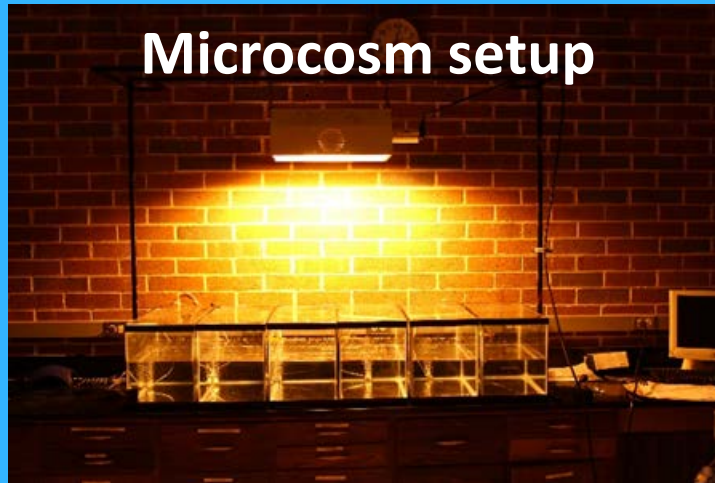
- Benthotorch®: no significant relationship with Taxonomist
- Using light adjustment & continuous measurements correlated better with laboratory results
- Variance within Benthotorch® readings increased with increasing chlorophyll a
- Improved accuracy with non-filamentous

Laboratory Methods

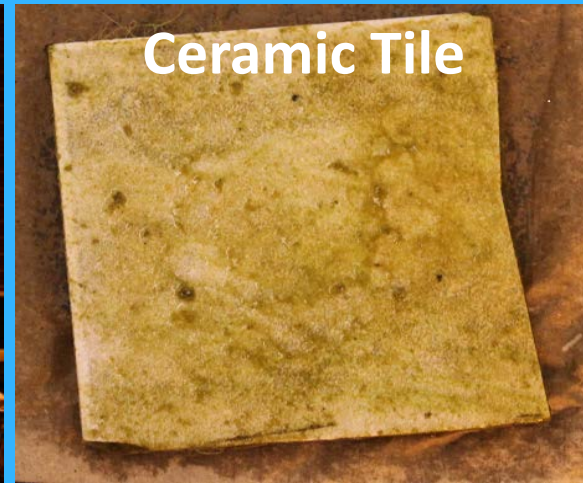
Sample collection



Microcosm setup



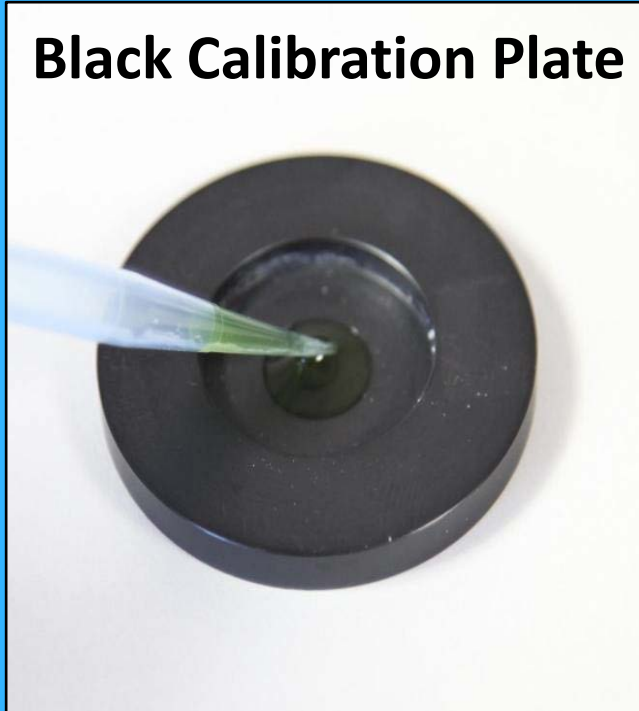
Ceramic Tile



**Benthometer
Reading on Tile**



Black Calibration Plate



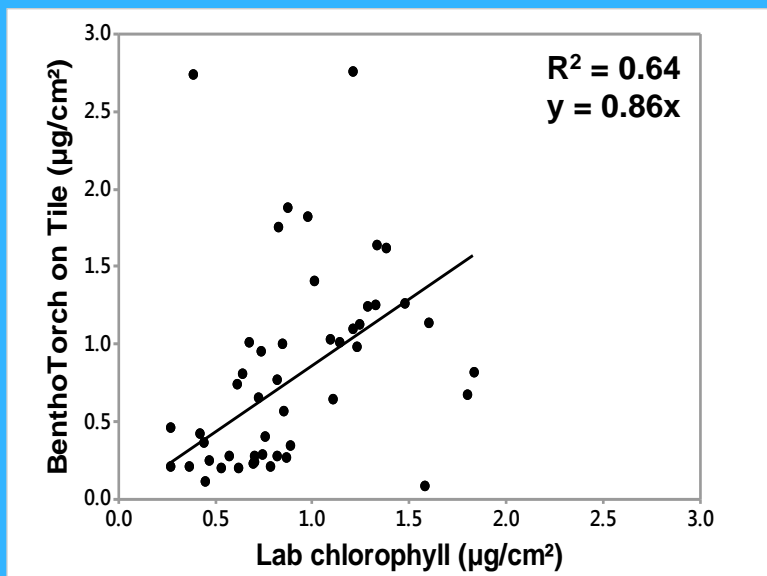
Spectrophotometer



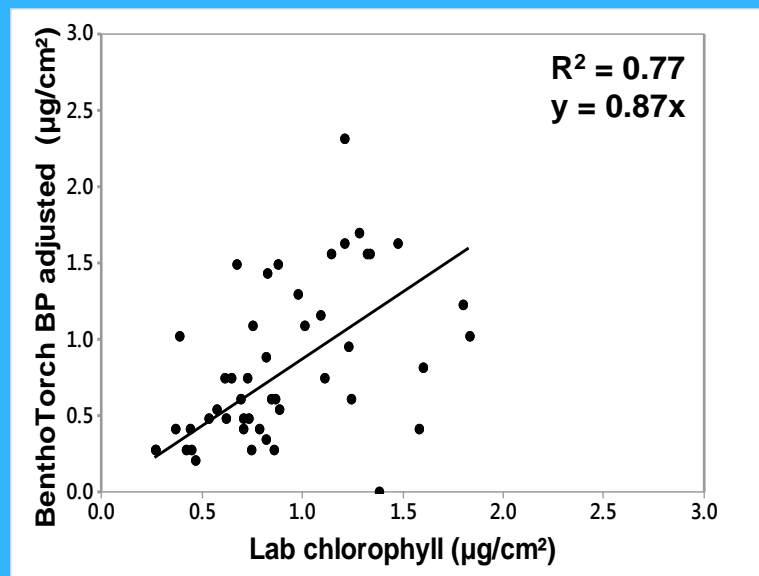
Results: Laboratory 2015-2016

BenthoTorch® Comparison with Lab Extracted Chlorophyll

In Situ Tile



Black Calibration Plate



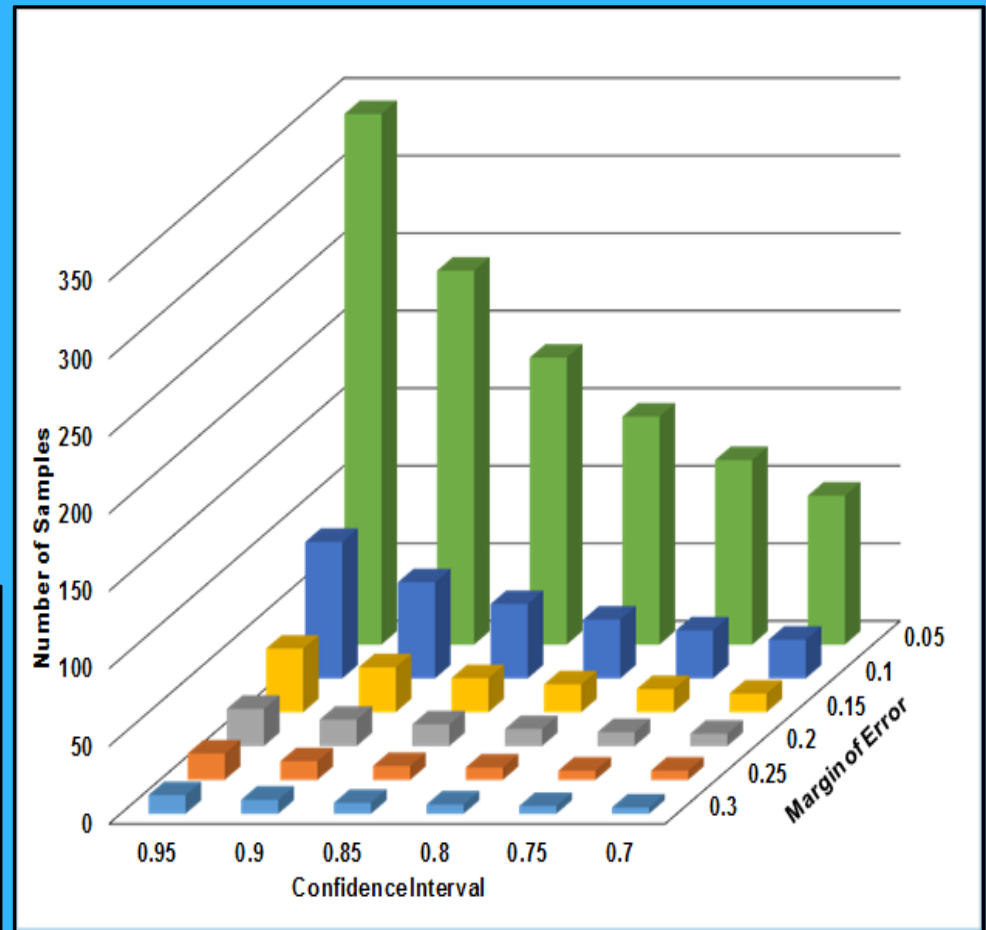
- Significant ($\alpha=0.05$) and reasonable regression equations
- Mean BenthoTorch® vs lab chlorophyll *a* not significantly different (paired t-test, $\alpha=0.05$)

Laboratory Study Conclusions

- BenthosTorch® *In Situ* Laboratory Tiles
 - Compares favorably in controlled environment with low chlorophyll *a* concentrations
- Modified Black Calibration Plate method looks promising for field conditions
 - Needs additional testing at sites with higher periphyton density
- BenthosTorch® likely a good tool to detect trends in periphyton density

Further Studies

- Stream reach characterization methods must be developed
- Number of BenthosTorch® samples to equal one traditional sample with specific confidence and margin of error.
- RMSE from lab experiment used as a predicted standard deviation in the sample-size estimate equation used by Montana DEQ (2011)
- Graph based on RMSE of 0.47 $\mu\text{g}/\text{cm}^2$ chlorophyll *a*
- Needs validation in field



Questions?

